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Synthesis and Evaluation of Radioligands for Imaging Brain Nociceptin/Orphanin FQ Peptide (NOP) Receptors with Positron Emission Tomography

Victor W. Pike,*[,†] Karen S. Rash,[‡] Zhaogen Chen,[‡] Concepción Pedregal,[§] Michael A. Statnick,[‡] Yasuyuki Kimura,[†] Jinsoo Hong,[†] Sami S. Zoghbi,[†] Masahiro Fujita,[†] Miguel A. Toledo,[§] Nuria Diaz,[§] Susan L. Gackenheimer,[‡] Johannes T. Tauscher,[‡] Vanessa N. Barth,[‡] and Robert B. Innis[†]

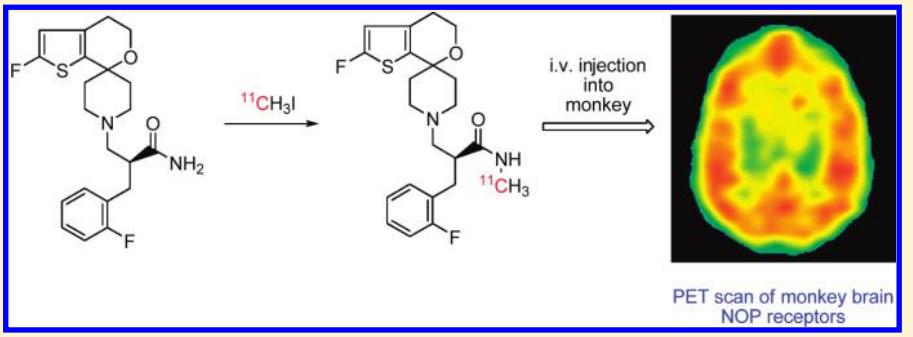
[†]Molecular Imaging Branch, National Institute of Mental Health, National Institutes of Health, Building 10, Room B3 C346A, 10 Center Drive, Bethesda, Maryland 20892, United States

[‡]Eli Lilly & Co., Lilly Research Laboratories, Indianapolis, Indiana 46285, United States

[§]Lilly S.A., Avenida de la Industria 30, 28108-Alcobendas, Madrid, Spain

Supporting Information

ABSTRACT:



Positron emission tomography (PET) coupled to an effective radioligand could provide an important tool for understanding possible links between neuropsychiatric disorders and brain NOP (nociceptin/orphanin FQ peptide) receptors. We sought to develop such a PET radioligand. High-affinity NOP ligands were synthesized based on a 3-(2′-fluoro-4′,5′-dihydrospiro[piperidine-4,7′-thieno[2,3-*c*]pyran]-1-yl)-2(2-halobenzyl)-*N*-alkylpropanamide scaffold and from experimental screens in rats, with ex vivo LC-MS/MS measures, three ligands were identified for labeling with carbon-11 and evaluation with PET in monkey. Each ligand was labeled by ^{11}C-methylation of an *N*-desmethyl precursor and studied in monkey under baseline and NOP receptor-preblock conditions. The three radioligands, [^{11}C](***S***)-**10a**−**c**, gave similar results. Baseline scans showed high entry of radioactivity into the brain to give a distribution reflecting that expected for NOP receptors. Preblock experiments showed high early peak levels of brain radioactivity, which rapidly declined to a much lower level than seen in baseline scans, thereby indicating a high level of receptor-specific binding in baseline experiments. Overall, [^{11}C](***S***)-**10c** showed the most favorable receptor-specific signal and kinetics and is now selected for evaluation in human subjects.

INTRODUCTION

Nociceptin, also known as orphanin FQ, is the most recently discovered peptide member of the endogenous opioid family.[1,2] Nociceptin acts on specific G-protein coupled receptors that are linked to the cAMP cascade and to voltage-gated Ca^{2+} and K^{+} channels.[3] These receptors were formerly known as ORL-1 (opioid receptor-like receptor-1) and are now called NOP (nociceptin opioid peptide) receptors. They are present in various organs, including the brain, and have roles in many normal physiological responses such as cognition, locomotion, and neuroendocrine control.[3] Additionally, many neuropsychiatric disorders are possibly linked to the NOP receptor, including pain, anxiety, depression, anorexia, obesity, and drug abuse.[4,5] Accordingly, the brain NOP receptor is a target not only for biomedical investigation but also for drug therapies.[4]

Positron emission tomography (PET),[6] in tandem with selective radioactive probes (radioligands), is a molecular imaging technique with a unique capability for investigating the living human brain and in particular for investigating specific proteins as possible players in pathophysiology[7] or as targets for therapeutic interventions.[8] Effective PET radioligands now exist for a wide range of targets, selected from among brain enzymes, transporters, plaque, ion channels, and neurotransmitter receptors.[9]

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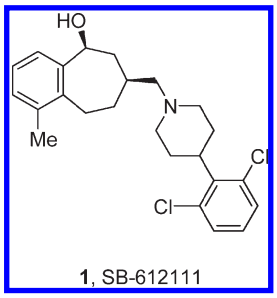
Chart 1. Structure of 1



1, SB-612111

Even so, many specific molecular targets of potential interest, including the NOP receptor, still lack a specific radioligand for investigation with PET. PET coupled to an effective NOP receptor radioligand could be an important tool for furthering the understanding of neuropsychiatric disorders in which NOP receptors are implicated and also for the development of new drugs that may target NOP receptors in the treatment of such disorders. We therefore sought to develop a radioligand for imaging brain NOP receptors with PET.

Effective PET radioligands for imaging in brain must display an array of favorable attributes, including amenability to labeling with a short-lived positron-emitter such as carbon-11 ($t_{1/2}$ = 20.4 min) or fluorine-18 ($t_{1/2}$ = 109.7 min), ability to cross the blood−brain barrier, high affinity and selectivity for binding to the target protein, relatively low nonspecific binding, and absence of troublesome brain-penetrant radiometabolites.[10−13] The difficult challenge of meeting such a combination of requirements in any proposed PET radioligand is a major reason for the current prevalence of nonimageable protein targets. A further important consideration is that the target protein should be present in adequate concentration for potential imaging. As a guideline the product of receptor density (B_{max}, nM) and ligand affinity ($1/K_D$, nM^{-1}) should exceed a value of 5 in target-rich regions of brain.

The distribution of NOP receptors in rat brain has been studied by autoradiography.[14−17] These studies show a widespread distribution of NOP receptors with, for example, relatively high levels in cortex, hypothalamus, and amygdala and low level in striatum. The density of NOP receptors in rat brain membranes was estimated at 237 fmol/mg protein (roughly equating to 24 nM in whole brain).[17] Data on the distribution and density of NOP receptors in the brains of higher species are sparse except for a single study on the macaque central nervous system.[18] This study found a distribution of NOP quite similar to that in rat but with some definite differences. Receptor densities of >6 fmol/mg (equating to >6 nM in whole brain) were found in receptor-rich regions such as cortex, caudate, and amygdala. Thus, prospective PET radioligands for brain NOP receptors should have high affinity, with a K_d value in the low nanomolar range.

The aforementioned autoradiographic studies were performed with peptide-like radioligands which would not be expected to enter the brain freely. Intense effort has been expended over recent years to discover nonpeptide small molecule ligands with high affinity as potential NOP receptor-targetted therapeutics.[19] A published example arising from this effort is SB-612111 (**1**; [(−)-*cis*-1-methyl-7-[[4-(2,6-dichlorophenyl)piperidin-1-yl]methyl]-6,7,8,9-tetrahydro-5*H*-benzocyclohepten-5-ol) (Chart 1).[20,21] From our own medicinal chemistry effort, we discovered a new series of high-affinity small molecule NOP receptor ligands, based on a 3-(2′-fluoro-4′,5′-dihydrospiro[piperidine-4,7′-thieno[2,3-*c*]pyran]-1-yl)-2(2-halobenzyl)-*N*-alkylpropanamide scaffold, which we disclose here. In this study, we explored this chemotype for the development of a PET radioligand for the NOP receptor, initially through screening of nonradioactive ligands in rat with ex vivo LC-MS/MS measures[22,23] and then through evaluation of selected ^{11}C-labeled ligands in rhesus monkey with PET. A promising NOP radioligand, [^{11}C](*S*)-**10c**, was found, which now merits further evaluation in human subjects.

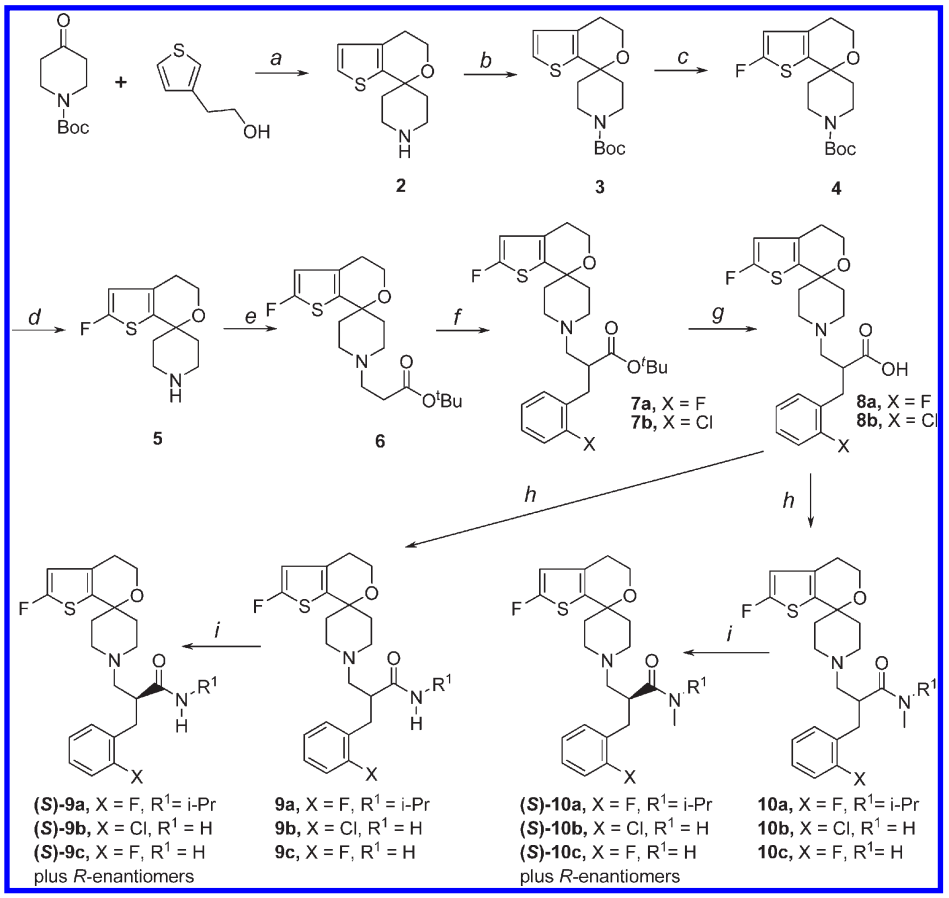
RESULTS AND DISCUSSION

Chemistry. The syntheses of the potential NOP ligands are outlined in Scheme 1. The spiro[4,5-dihydrothieno[2,3-*c*]pyran-7,4′-piperidine] **2** was synthesized from 2-(3-thienyl)ethanol by treatment of *t*-butylcarbonyl-protected (Boc-protected) piperidin-4-one in the presence of trifluoroacetic acid (TFA) in good yield. After the piperidine nitrogen was protected with Boc, fluorine was selectively introduced at the C2 position of the thienopyranyl ring via metalation with *n*-BuLi and subsequent reaction with the fluorination reagent *N*-fluoro-*N*-(phenylsulfonyl)-benzenesulfonamide (NFSI) in 40−50% yield. Michael addition between the piperidine intermediate **5** and *t*-butyl acrylate under mild conditions gave the common intermediate propanoate **6** in 65% yield. α-Alkylation of propanoate **6** with either 2-fluorobenzyl bromide or 2-chloro-benzyl bromide was achieved with $LiN(TMS)_2$ as base. Ester hydrolysis of compounds **7a** and **7b**, followed by conventional amide formation with the appropriate amine afforded the final amides **9a**−**c** and **10a**−**c**, which were each separated into enantiomers with chiral HPLC. The biologically active enantiomer of **9c** was determined by single-crystal X-ray crystallography to have *S*-configuration (see Supporting Information). The *R*-enantiomer of **9c** has markedly lower affinity (Table 1). The biologically less active enantiomer of **9a** was also determined by single-crystal X-ray crystallography to have *R*-configuration (see Supporting Information). We therefore reasoned that the higher affinity enantiomers of the NOP ligands **9a**, **9b**, and **10a**−**c** also have *S*-configuration

To assess the stereochemical stability of the chiral center in the new ligands, as well as explore possible future conditions of radiolabeling with [^{11}C]methyl iodide, compound (*S*)-**9a** was treated with sodium hydride in dimethyl sulfoxide (DMSO) solution at ambient temperature and then with methyl iodide. Chiral HPLC showed that this gave a single enantiomer of the *N*-methylated product **10a**, plus unchanged starting material, (*S*)-**9a**. Therefore, these basic alkylation conditions do not cause significant racemization and the obtained product also has *S*-configuration (i.e., the product is (*S*)-**10a**).

Ligand Lipophilicity. Consideration of ligand lipophilicity is important in the development of PET radioligands. Moderate lipophilicity is usually sought to promote measurable plasma free fraction (f_P) and adequate brain entry, avoid excessive nonspecific binding to brain tissue, and avoid the generation of lipophilic brain-penetrant radiometabolites.[10,13,24] The lipophilicity (log *D*) of (*S*)-**10c** was measured to be 3.41 by a radiometric method and 3.50 by a shake-flask method (Table 1). This lipophilicity is within a range that is considered favorable for achieving adequate brain entry of a PET radioligand from blood without incurring excessive nonspecific binding to brain tissue.[10,13,24] For ligands **10a**−**c**, the calculated lipophilicities (cLogD values at pH 7.4) were only slightly lower than those measured, so indicating that

Scheme 1. Synthesis of NOP Ligands 9a−c and 10a−c and Their Enantiomers



Reagents and conditions: (a) (1) TFA/CH_2Cl_2, rt, (2) 10% NaOH aq; (b) di-*t*-butyl dicarbonate, TEA, CH_2Cl_2; (c) *n*-BuLi, −78 °C, NFSI, THF; (d) 4 M HCl in dioxane, CH_2Cl_2, rt; (e) *t*-butyl acrylate, TEA,THF, reflux; (f) (1) LiN(TMS)$_2$, 1 M THF, 1,3-dimethyl-tetrahydropyrimidin-2(1*H*)-one, THF, −78 °C, (2) 2-F-benzyl bromide or 2-Cl-benzyl bromide; (g) TFA/CH_2Cl_2, rt; (h) EDCI, HOBt, DIPEA, CH_2Cl_2, NH_4Cl or appropriate amine; (i) chiral HPLC separation.

Table 1. Receptor Binding Affinities, Antagonist Activities, and Lipophilicities of NOP Ligands

ligand	NOP receptor binding[a] K_i (nM)	NOP antagonist activity[a] K_b (nM)	cLogD[b]
1	0.253 ± 0.151	0.258 ± 0.129	4.98
(*S*)-**9a**	0.231 ± 0.050	0.228 ± 0.050	4.08
(*R*)-**9a**	4.51 ± 1.05	5.64 ± 1.05	4.08
(*S*)-**9b**	0.101 ± 0.029	0.110 ± 0.021	3.41
(*R*)-**9b**	17.1 ± 2.54	7.36 ± 0.019	3.41
(*S*)-**9c**	0.106 ± 0.049	0.173 ± 0.084	2.95
(*R*)-**9c**	5.61 ± 2.74	8.65 ± 5.19	2.95
(*S*)-**10a**	0.137 ± 0.048	0.141 ± 0.032	4.39
(*R*)-**10a**	10.2 ± 2.85	15.8 ± 3.48	4.39, (4.47)[c]
(*S*)-**10b**	0.090 ± 0.016	0.099 ± 0.048	3.68
(*R*)-**10b**	9.08 ± 1.04	5.79 ± 1.05	3.68, (3.90)[c]
(*S*)-**10c**	0.150 ± 0.062	0.069 ± 0.015*	3.27, (3.50)[c], (3.41 ± 0.07, *n* = 6)[d]
(*R*)-**10c**	5.32 ± 1.03	7.92 ± 1.05	3.27

[a] Assays were performed in triplicate except for that with an asterisk (n = 2). Data are mean ± SE. [^{3}H]Nociceptin was used as reference NOP radioligand in a binding assay using human recombinant NOP receptors expressed in CHO cells. Antagonist activity was determined in an assay of receptor-mediated G-protein activation using [^{35}S]GTPγS and membranes expressing cloned human NOP receptors. See Materials and Methods for assay details. [b] Computed with Pallas software. Values in parentheses are measured values. [c] log *D* measured with nonradioactive compound and shake-flask method. [d] log *D* value (mean ± SD) measured with [^{11}C](*S*)-**10c**.

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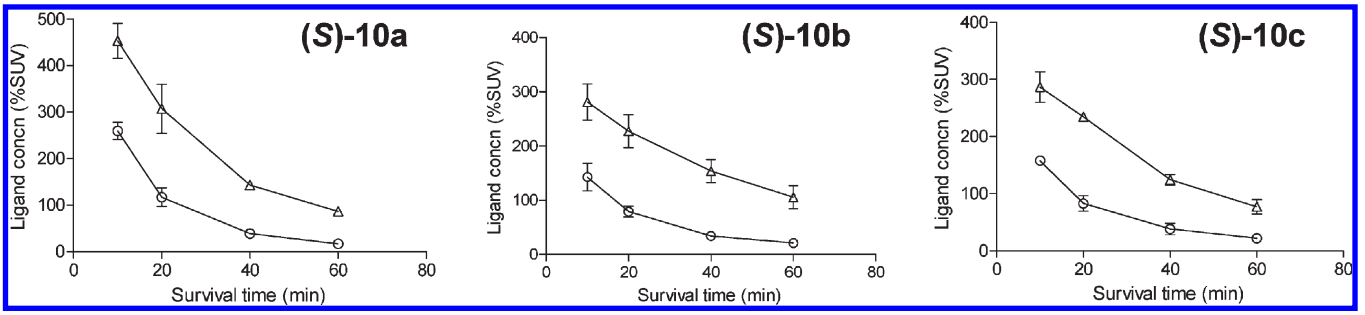


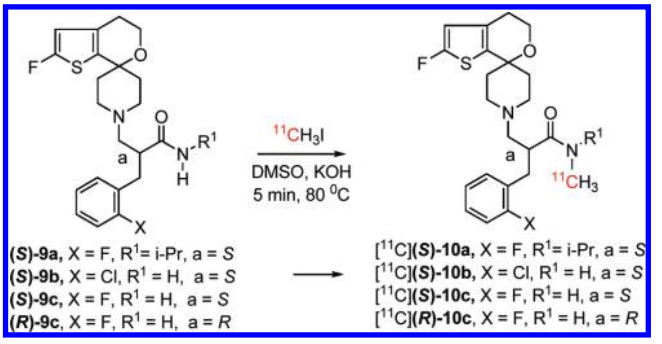
Figure 1. Time-courses of normalized ligand concentrations (%SUV) in rat hypothalamus (△) and striatum (○) after iv injection of nonradioactive NOP ligand (*S*)-**10a**, (*S*)-**10b** or (*S*)-**10c** (3 μg/kg, iv), measured ex vivo with LC-MS/MS. Error bars are SD (where not visible they are within the symbol size).

the computation on this type of structure was quite accurate. As expected, from the extra presence of an iso-propyl group, the cLogD and measured log *D* values of ligand **10a** were appreciably higher than those of **10c**, as where those of **10b** in which a fluoro substituent of **10c** is replaced with a chloro substituent. The corresponding desmethyl compounds were computed to have less lipophilicity, as would also be expected.

Ligand Pharmacology. Ligands (*S*)-**10a**−**c** were found to have high binding affinity to human recombinant NOP receptors as represented by sub-nM K_i values in a binding assay with [^{3}H]nociceptin as reference NOP radioligand (Table 1). In a functional assay of receptor-mediated G-protein activation with [^{35}S]GTPγS, these ligands were each found to be potent antagonists at NOP receptors with sub-nM K_b values (Table 1). Thus, (*S*)-**10a**−**c** meet the high-affinity criterion previously mentioned for prospective NOP PET radioligands. The *R*-enantiomers were found to have substantially lower binding affinities and antagonist activities. Eudismic ratios ranged from 35 to 115. The enantiomers of the corresponding *N*-desmethyl ligands, **9a**−**c**, showed a similar pattern of binding affinity and antagonist activity and displayed eudismic ratios ranging from 20 to 169 (Table 1).

Studies in Rats. Initially, we applied our previously published in vivo/ex vivo LC-MS/MS method to characterize NOP ligands as potential tracers in rats because this method avoids the need to use radiolabels.[22,23] Essentially, a low amount of nonradioactive ligand was injected intravenously into a group of rats, and after a set time the animals were sacrificed. Brain tissue regions, corresponding to high NOP and low NOP receptor expression, namely hypothalamus and striatum, respectively,[15,16] were dissected out and their concentrations of ligand measured with LC-MS/MS. As a result, we identified three high-affinity ligands, (*S*)-**10a**−**c**, with potential for labeling to produce effective PET radioligands for brain NOP receptors. After intravenous administration into nonanesthetized rat, all three ligands showed quite fast kinetics with rapid and high rat brain uptake and washout occurring within 1 h, a time-span consistent with the prospective use of short-lived carbon-11 as a radiolabel (Figure 1). Each ligand showed high brain uptake with ligand (*S*)-**10a** having slightly higher uptake (453% SUV at 10 min) than (*S*)-**10b** or (*S*)-**10c** (each ∼280% SUV). All three ligands robustly demonstrated about 3-fold higher uptake in NOP-rich hypothalamus than in NOP-poor striatum by 40 min after injection, thereby indicating a sizable amount of NOP receptor-specific binding. Ligand (*S*)-**10b** had the largest ratio of NOP receptor-specific to nonspecific binding (i.e., hypothalamus/striatum ligand concentration ratio minus 1 at 40 min), with a value of 3.5, whereas

Scheme 2. Radiolabeling of NOP Ligands



ligands (*S*)-**10a** and (*S*)-**10c** had ratios of 2.7 and 2.3, respectively. These ratios are in fact underestimates of specific to nonspecific binding in hypothalamus because rat striatum is not entirely devoid of NOP receptors.[15,16]

Radioligand Syntheses. Ligands (*S*)-**10a**−**c** and (*R*)-**10c** were rapidly labeled with carbon-11 by treating their respective *N*-desmethyl precursors, (*S*)-**9a**−**c** and (*R*)-**9c**, with [^{11}C]methyl iodide in the presence of KOH as base (Scheme 2). The radioligands were isolated with reverse phase HPLC in useful activities (mean yields for n = 3 were 47, 75, and 85 mCi for [^{11}C](*S*)-**10a**−**c**, respectively), and in high radiochemical purities (>99%) and specific radioactivities (mean values for n = 3 were, 7.8, 5.1, 3.6 Ci/μmol at end of synthesis for [^{11}C](*S*)-**10a**−**c**, respectively). Chiral HPLC analyses of the radioligands verified almost complete absence of racemization in the labeling procedure (see Supporting Information for chiral HPLC analysis of [^{11}C](*S*)-**10c**). Each radioligand was readily formulated in ethanol (10% v/v)/saline for intravenous administration into monkey. Each was radiochemically stable in formulation medium. The radiosyntheses times were about 45 min from the end of radionuclide production.

PET Imaging in Monkeys. After intravenous administration into monkey, each high-affinity radioligand, [^{11}C](*S*)-**10a**−**c**, showed similar brain radioactivity kinetics, characterized by rapid high uptake into brain followed by progressive washout of radioactivity from all examined regions over the duration of the scanning session (either 90 or 120 min) (Figure 2). Cerebellum showed highest initial uptake and fastest washout of radioactivity in each case, possibly due to greater regional blood flow in this region of brain during anesthesia. The rank order of initial radioactivity uptake was [^{11}C](*S*)-**10c** > [^{11}C](*S*)-**10b** > [^{11}C](*S*)-**10a**. [^{11}C](*S*)-**10c** also gave the greatest separation of time−activity

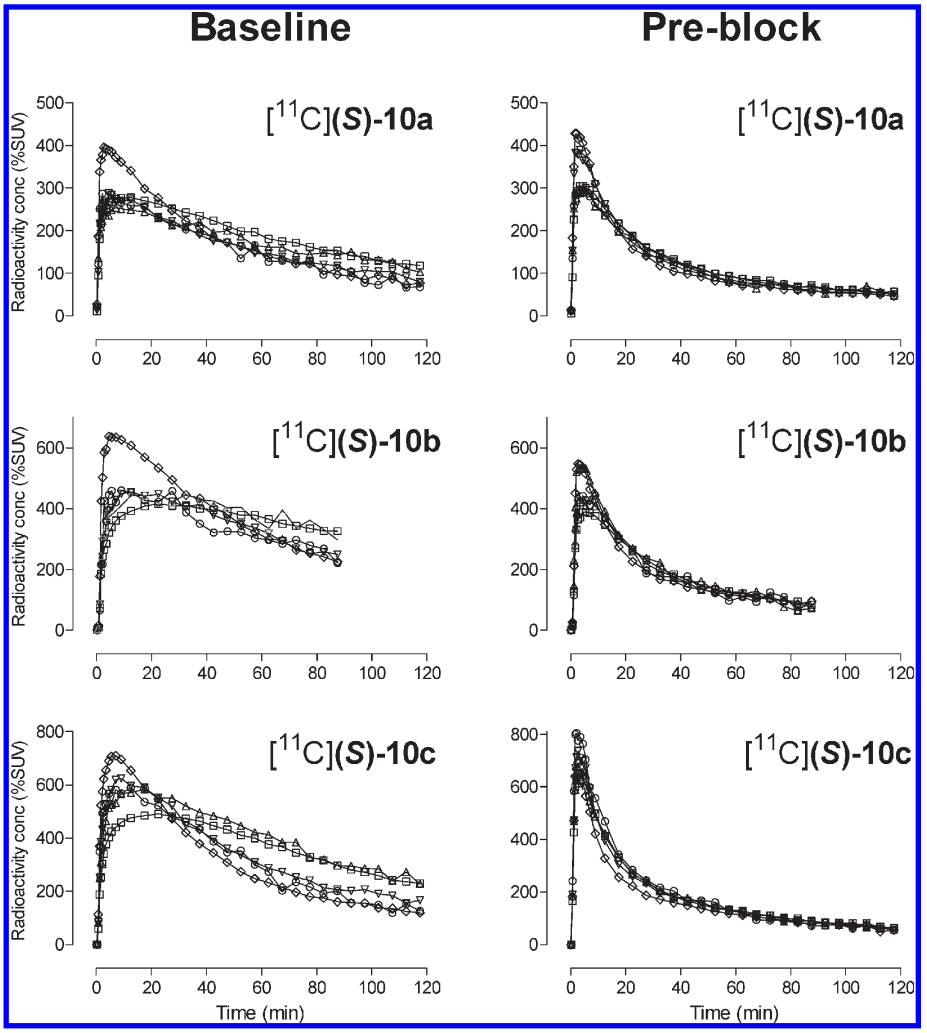


Figure 2. Brain region time−activity curves in monkey after injection of radioligands [^{11}C](*S*)-**10a**−**c** in baseline and in preblock experiments, in which NOP preblocker **1** (2 mg/kg, iv) was given at 10 min before radioligand. For each radioligand, the baseline and preblock experiments were conducted about 3 h apart in the same monkey. Key: hypothalamus (○), cerebellum (◇), caudate (▽), amygdala (△), lateral temporal cortex (□). Time−activity curves for other examined regions as listed in the experimental were generally similar to the noncerebellar curves.

curves across brain regions. Compound **1** is a highly potent (Table 1) and selective antagonist for NOP receptors, showing more than 1000-fold lower affinity for classical opioid receptors.[21] In corresponding experiments in which brain NOP receptors were preblocked by administration of **1**, brain radioactivity kinetics were altered in a consistent manner (Figure 2). Thus, in each monkey, maximal initial radioactivity uptake was again high but was followed by a much faster washout of radioactivity from all examined brain regions than in the corresponding baseline scans and with very little separation in the time−activity curves for noncerebellar regions. Such differences in kinetics imply the presence of a significant proportion of NOP receptor-specific binding of the radioligand in the baseline experiments. Major differences were also clearly apparent in the PET images obtained under baseline and preblock conditions. Thus, the baseline experiments showing a regional distribution of radioactivity reflecting the distribution of NOP receptors expected from the autoradiographic study in macaque brain,[18] with, for example, high uptake in caudate and relatively low uptake in hypothalamus, the reverse of relative levels in rat.[15,16] The preblock experiments in monkey showed low and uniform distributions of radioactivity consistent with nonspecific binding of radioligand and/or radiometabolites. These image differences are exemplified in Figure 3 for [^{11}C](*S*)-**10c**.

Comparison of the areas-under-the-curves (AUCs) between baseline and preblock PET experiments provided further evidence for an appreciable proportion of NOP receptor-specific binding under baseline conditions. Thus, for each radioligand, the brain-averaged AUCs (%SUV × min) between 0 and 90 min or between 30 and 90 min were appreciably greater in the baseline experiments than in the preblock experiments. From these data, crude estimates of ratios of NOP receptor-specific binding to nonspecific binding were derived as $[(AUC_{baseline}/AUC_{preblock}) - 1]$ for the time intervals 0−90 and 30−90 min after radioligand injection. These estimates (Table 2) confirm that each radioligand produced a high proportion of NOP receptor-specific binding in baseline PET scans, with radioligands [^{11}C](*S*)-**10b** and [^{11}C](*S*)-**10c** showing appreciably higher proportions than [^{11}C](*S*)-**10a**. (A full biomathematical compartmental analysis of monkey PET scans to derive accurate binding potentials is beyond the scope of this report and will be published subsequently.)

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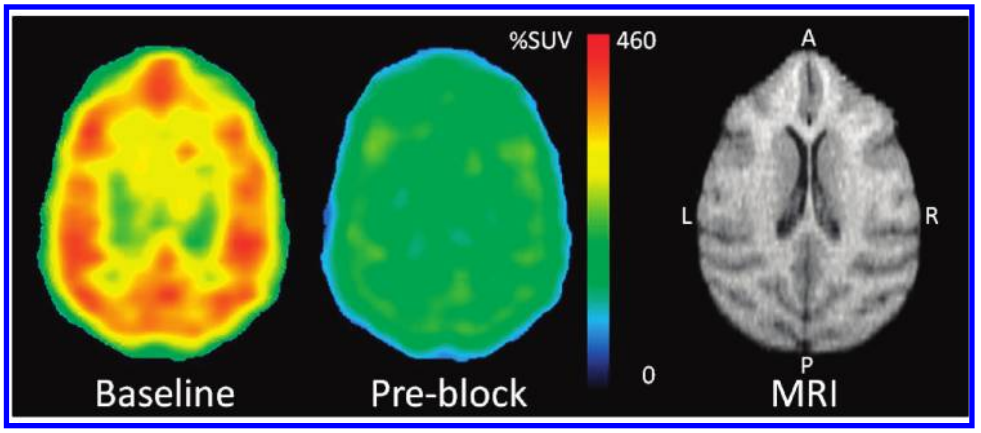

Figure 3. Horizontal PET images of monkey brain at the level of the caudate acquired between 0 and 100 min after intravenous injection of [^{11}C](*S*)-**10c** under baseline and preblock conditions. The corresponding magnetic resonance imaging (MRI) scan shows PET scan orientation and anatomy.

Table 2. NOP Receptor-Specific to Nonspecific Binding Ratios for Radioligands [^{11}C](*S*)-10a−c, in Monkey Estimated from (a) Brain-Averaged AUCs over 0−90 min (Method A), or (b) over 30−90 min (Method B)

		specific to nonspecific ratio[a]	
ligand	*n*	method A	method B
[^{11}C](*S*)-**10a**	2	0.19, 0.44	0.53, 0.84
[^{11}C](*S*)-**10b**	3	0.66 ± 0.26	1.31 ± 0.35
[^{11}C](*S*)-**10c**	5	0.62 ± 0.36	1.28 ± 0.70

[a] The NOP receptor-specific to nonspecific binding ratio was calculated as $[(AUC_{baseline}/AUC_{preblock}) - 1]$, where the AUCs are measured over the same time interval.

Radioligand [^{11}C](*S*)-**10c** was selected for deeper investigation because of its high brain uptake and apparently high ratio of NOP receptor-specific to nonspecific binding. Administration of the selective NOP ligand **1** to monkey at 40 min after [^{11}C](*S*)-**10c** resulted in a faster washout of radioactivity from all brain regions down to a low level, so demonstrating the reversibility of the specific binding of [^{11}C](*S*)-**10c** to NOP receptors (Figure S4, Supporting Information). The opiate receptor antagonist naloxone is effective at displacing opiate receptor radioligands from primate brain.[25,26] In another monkey PET experiment, administration of a high dose of naloxone at 37 min after [^{11}C](*S*)-**10c** had no effect on the washout of radioactivity from brain, thereby showing that this radioligand does not bind significantly to opiate receptors (Figure S5, Supporting Information). Furthermore, baseline and preblock experiments in a single monkey with [^{11}C](*R*)-**10c**, the lower affinity enantiomer of [^{11}C](*S*)-**10c**, provided no evidence of receptor-specific binding because they showed similar fast kinetics in all brain regions with no separation of regional time−activity curves (Figure S6, Supporting Information). Maximal brain radioactivity uptake was in fact higher in the preblock experiment.

Emergence of Radiometabolites of [^{11}C](*S*)-10c in Monkey Plasma in Vivo. Generally, PET radioligands are appreciably metabolized over the short time-span of a PET scanning session. Detailed quantitative analysis of radioligand behavior in vivo to derive important output measures such as binding potential or volume of distribution may require accurate measurement of f_P and also the determination of how the amount of unmetabolized radioligand in plasma changes over the full duration of a scanning session (the radioligand arterial input function). Generally, PET radioligands that produce appreciable amounts of brain-penetrant radiometabolites are difficult to quantify.

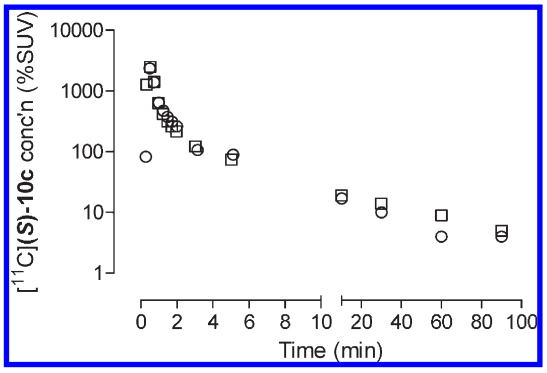

Figure 4. Time-course of concentration of unmetabolized [^{11}C](*S*)-**10c** in plasma after intravenous injection into monkey under baseline (○) and preblocked conditions (□).

[^{11}C](*S*)-**10c** was found to be radiochemically stable (98.9 ± 0.4% unchanged, $n = 4$) in sodium phosphate buffer (0.15 M, pH 7.4) for 2.5 h and also stable in monkey blood and plasma for 0.5 h at room temperature [100.4 ± 0.01% ($n = 6$) and 99.9 ± 0.01% ($n = 6$)]. The f_P value of [^{11}C](*S*)-**10c** in monkey blood in vitro was quite low (11.3 ± 1.1%; $n = 6$, 3 samples from each of two monkeys) but readily measurable with good accuracy. The f_P value in human pooled plasma was similar to that in the monkey (11.0 ± 0.7%; three samples from each of two subjects). The values are within the range expected for the lipophilicity of (*S*)-**10c** (log $D = 3.41$).[27]

After administration of [^{11}C](*S*)-**10c** into monkey, both the total radioactivity in plasma and the parent radioligand component reduced rapidly (Figure 4). Recoveries of radioactivity from plasma into supernatant acetonitrile for HPLC analysis were 94.9 ± 3.3% ($n = 106$). HPLC analyses of plasma from monkeys studied at baseline revealed [^{11}C](*S*)-**10c** eluting at 4.4 ± 0.8 min ($n = 100$) and three less lipophilic radiometabolites eluting at 1.7 ± 0.4, 2.2 ± 0.5 and 2.7 ± 0.6 min ($n = 100$), respectively. [^{11}C](*S*)-**10c** was rapidly metabolized to a virtually equal extent in both baseline and preblock experiments in the same monkeys, respectively (Figure 5). The identities of the three radiometabolites are currently unknown, as is their ability to penetrate the

blood−brain barrier. Their lower lipophilicities relative to that of [^{11}C](*S*)-**10c**, as deduced from their shorter retention times on reverse phase HPLC, might render them less brain-penetrant and untroublesome to quantitative analysis. As shown in Figure 2, treatment of monkey with the NOP antagonist **1** before [^{11}C](*S*)-**10c** resulted in a continuous decline of radioactivity in all brain regions to a low level, representing about 15% of the early peak value at the end of the scanning session (~120 min). Radiometabolites are therefore unlikely to be entering brain to a troublesome extent. (Our detailed kinetic analysis of [^{11}C](*S*)-**10c** in monkey, to be published elsewhere, show that stable volumes of distribution values are established rapidly and therefore that radiometabolites do not enter brain to be problematic to receptor quantification.)

Test of Radioligand Selectivity by in Vitro Autoradiography of Wild-Type and NOP Knockout Mouse Brain. Autoradiographs of post mortem wild-type mouse brain slices with [^{3}H$_3$](*S*)-**10c** showed a discrete distribution of radioligand uptake (Figure 6), and they were very comparable with those previously reported for rat using [^{3}H]nociceptin[15] or other peptide-like NOP receptor radioligands.[14,16,17] [^{3}H$_3$](*S*)-**10c** gave only low nonspecific binding. Autoradiographs in NOP receptor knockout mice showed only a very low uniform distribution of [^{3}H](*S*)-**10c**, so demonstrating the specificity of the binding of this radioligand to NOP receptors in wild-type mouse brain.

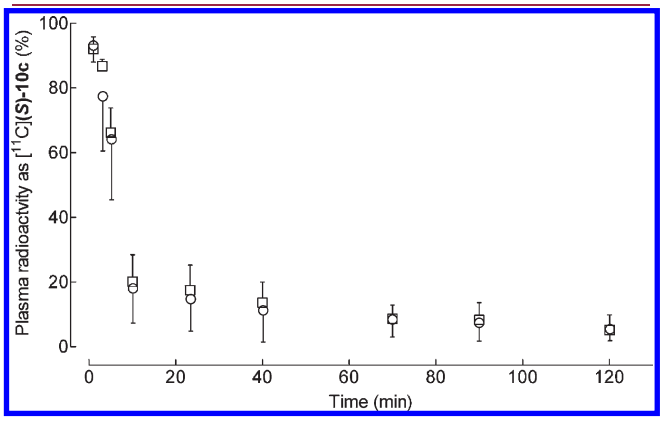

Figure 5. Percentage of radioactivity in plasma represented by [^{11}C](*S*)-**10c** during PET scanning of three monkeys under baseline (○) and preblock conditions (□). Error bars are SD values.

SUMMARY

The 3-(2′-fluoro-4′,5′-dihydrospiro[piperidine-4,7′-thieno-[2,3-*c*]pyran]-1-yl)-2(2-halobenzyl)-*N*-alkylpropanamides represent a new chemotype of high-affinity NOP receptor ligand. This study shows that this chemotype is suitable for the development of prospective NOP receptor PET radioligands, of which [^{11}C](*S*)-**10c**, on account of its good brain entry, favorable kinetics, and strong receptor-specific signal in rat and monkey in vivo, is particularly promising for further evaluation in human subjects.

EXPERIMENTAL SECTION

Materials and Methods. All reagents used were obtained from commercial sources (Sigma-Aldrich, unless otherwise stated). All solvents were of an analytical grade. The NOP ligand, **1**,[20,21] used as a NOP blocking or NOP ligand displacing compound, was synthesized at Eli Lilly and Company. [^{3}H](*S*)-**10c** was prepared by treating the *N*-desmethyl compound (*S*)-**9c** with [^{3}H$_3$]methyl iodide and obtained in >99% radiochemical purity, as determined with HPLC on a reverse phase C18 column (5 μm; 4.6 mm × 150 mm) eluted with a mixture of 0.05% TFA (A) and MeCN (B) at 1 mL/min with B increased from 30% v/v to 90% at 10 min and returned to 30% at 12 min (t_R = 7.89 min). The specific activity of [^{3}H](*S*)-**10c** was found to be 53 Ci/mmol with mass spectrometry. [^{35}S]GTPγS was obtained commercially.

Isolation of compounds by column chromatography was performed on silica gel columns (SP1a HPFC system, Biotage Inc., Charlottesville, VA). ^{1}H NMR (300 MHz) spectra were acquired at 20 °C on a System 300 instrument (Varian; Palo Alto, CA). Abbreviations s, d, dd, ddd, m, and br denote singlet, doublet, double doublet, double double doublet, multiplet, and broad, respectively.

LC-MS was performed with an 1100 Series LC-MSD single quadrupole instrument (Agilent; Santa Clara, CA) with an ESI interface. Samples were analyzed by method 1, 2, or 3. Method 1 used a heated (50 °C) Gemini C18 column (3.0 μm; 2.0 mm × 50 mm; Phenomenex, Torrance, CA) eluted at 1 mL/min with a gradient of A (H_2O containing 0.01%TFA) and B (MeCN containing 0.01% TFA) with B increased linearly from 5% to 100% (v/v) over 7 min and then held for 1 min. Method 2 analyses used a heated (50 °C) Gemini C18 column (3.0 μm;

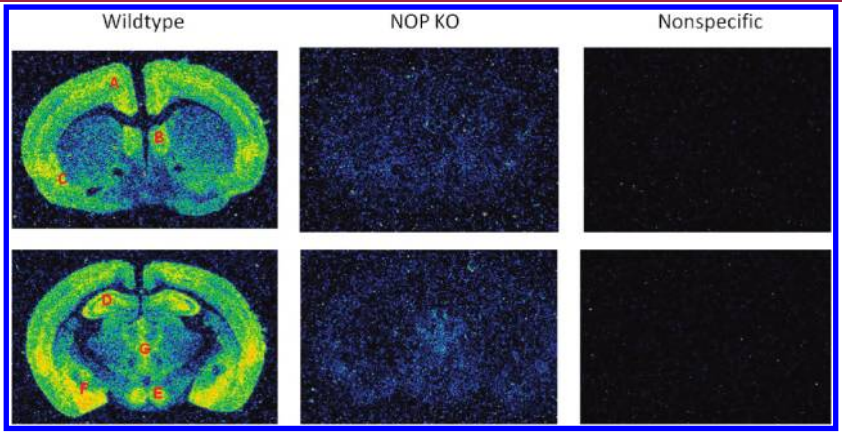

Figure 6. Autoradiographs of binding of [^{3}H$_3$](*S*)-**10c** (0.6 nM) to 12 μm coronal sections of wild-type 129/S6 and NOP knockout mouse brain. Nonspecific binding was determined in the presence of 10 μM **1** (from an adjacent section from wild-type mice). Wild-type 129/S6 mice exhibited high binding in numerous cortical regions (A), medial septum (B), dorsal endopiriform nucleus (C), hippocampus (D), vetromedial hypothalamic nucleus (E), amygdala (F), and central thalamus (G). No appreciable specific binding was observed in equivalent sections of NOP knockout mouse brain.

2.0 mm × 50 mm) eluted at 1 mL/min with a gradient of A (aq NH_4HCO_3; 10 mM) and B (MeCN), with B increased linearly from 10% to 100% (v/v) over 7 min and then held for 1 min. Method 3 used an XBridge C18 column (3.5 μm; 2.1 mm × 50 mm; Waters, Milford, MA) eluted at 1 mL/min with a gradient of A (aq NH_4HCO_3; 10 mM, pH 9) and B (MeCN), with B increased linearly from 10% to 100% (v/v) over 7 min and then held for 1 min. Ions between *m/z* 80 and 800 were captured after electrospray ionization of the eluted test sample. The purities of synthesized ligands were all found to be ≥95%, by methods 1, 2, or 3 as specified later.

High-resolution mass spectrometry (HRMS) was performed on an TOF/Q-TOF instrument (Agilent). Data were acquired in dual ESI mode (+ and −) in the range of 95−1700 amu. The sample (~0.1 mg/mL in MeCN/H_2O, 1:1 v/v; 2 μL) was injected in loop mode onto a Zorbax SB-C8 column (3.5 μm; 3.0 mm × 150 mm; Agilent) eluted at 0.5 mL/min with a mixture of 0.1% formic acid in water (A) and 0.1% formic acid in MeCN (B) with B increasing from 5% to 100% over 15 min and then held at 100% for 8 min. Eluate was monitored with a diode array detector operating in the range 190−700 nm.

Melting points (mp) were determined on a DSC Q100 V9.8 instrument (Universal) with a ramp of 5 °C/min up to 300 °C, and were uncorrected.

Optical rotations were measured with a 341 polarimeter (Perkin-Elmer) at 20 °C and at 589 nm (sodium lamp).

γ-Radioactivity from carbon-11 was measured using a calibrated dose calibrator (Atomlab 300; Biodex Medical Systems; Shirley, NY). Radioactivity measurements were corrected for physical decay. All radiochemistry with carbon-11 was performed in a lead-shielded hot-cell for personnel protection from radiation.

Statistics. Values are given as means ± the standard deviation of the mean, unless otherwise stated.

Animals. Male Sprague−Dawley rats (Harlan Sprague−Dawley, Indianapolis, IN), weighing between 230 and 280 g, were used for in vivo experiments. All rats were housed in rooms using a 12 h light/dark cycle and had ad libitum access to normal rat chow and water until the beginning of the 3 h experimental protocol. All experiments with rats were performed in accord with the National Research Council Guide under protocols approved by the Animal Care and Use Committee of Eli Lilly and Company.

All rhesus (*Macaca mulatta*) monkeys were handled in accordance with the *Guide for the Care and Use of Laboratory Animals*[28] and the National Institute of Health Animal Care and Use Committee.

Chemistry. *Spiro[4,5-dihydrothieno[2,3-c]pyran-7,4′-piperidine] (**2**).* 2-(Thiophen-3-yl)ethanol (15.4 g, 120 mmol) was added to a solution of *N-t*-butoxycarbonyl-4-piperidone (20 g, 100 mmol) in CH_2Cl_2 (200 mL) at room temperature (rt). TFA (38 mL) was added dropwise to the mixture over 30 min, while the internal temperature was kept between 23 and 30 °C. The mixture was then stirred at rt for 20 h. Solvent was then evaporated off, and the residue was poured into NaOH (1M; 200 mL). The mixture was stirred for 15 min and the pH adjusted to about 11 with 50% aq NaOH. Then the mixture was extracted with CH_2Cl_2. The organic layers were separated, dried over Na_2SO_4, and evaporated. The crude residue was purified on silica gel (CH_2Cl_2/MeOH; 20:1 v/v) to afford 19.9 g (95%) of **2** as a solid. LC-MS (ESI) *m/z* 210 $(M + H)^+$, purity 100%, t_R = 1.68 min, method 2. ^{1}H NMR (300 MHz, $CDCl_3$): δ 7.14 (d, *J* = 4.8 Hz, 1H), 6.77 (d, *J* = 5.1 Hz, 1H), 3.93 (t, *J* = 5.4 Hz, 2H), 3.09 (td, *J* = 3.0 Hz, 12.3 Hz, 2H), 2.98−2.92 (m, 2H), 2.70 (t, *J* = 5.7 Hz, 2H), 2.01−1.99 (m, 2H), 1.86 (td, *J* = 4.5 Hz, 12.3 Hz, 2H).

*t-Butyl Spiro[4,5-dihydrothieno[2,3-c]pyran-7,4′-piperidine]-1-carboxylate (**3**).* Triethylamine (TEA, 0.97 g, 9.56 mmol) was added to a mixture of **2** (1.0 g, 4.78 mmol) and di-*t*-butyl dicarbonate (1.25 g, 5.74 mmol) in dry dichloromethane (20 mL) and stirred at rt for 5 h. The reaction mixture was then washed with 1 M HCl and brine, dried over Na_2SO_4, filtered, and concentrated in vacuo to afford **3** as a yellow oil (1.476 g, 100%). LC-MS (ESI): *m/z* 210 $(M + H - 100)^+$, purity 100%, t_R = 1.82 min, method 1. ^{1}H NMR (300 MHz, $CDCl_3$) δ 7.14 (d, *J* = 4.8 Hz, 1H), 6.77 (d, *J* = 5.1 Hz, 1H), 4.05−3.91 (m, 4H), 3.21−3.05 (m, 2H), 2.70 (t, *J* = 5.4 Hz, 2H), 2.00−1.96 (m, 2H), 1.80 (td, *J* = 5.1 Hz, 12.9 Hz, 2H), 1.52 (s, 9H).

*t-Butyl 2-Fluorospiro[4,5-dihydrothieno[2,3-c]pyran-7,4′-piperidine]-1′-carboxylate (**4**).* *n*-BuLi (1.6 M; 10.7 mmol) in hexane (6.7 mL) was added dropwise to a solution of **3** (1.102 g, 3.57 mmol) in dry THF (15 mL) at −78 °C under N_2. The reaction mixture was stirred at −78 °C for 1 h, and then a solution of NFSI (2.811 g, 8.93 mmol) in dry THF (30 mL) was added at −78 °C. The mixture was then warmed to rt and stirred overnight. The reaction was quenched with saturated NH_4Cl solution and the mixture extracted with EtOAc. The organic layer was washed with brine, dried over Na_2SO_4, and concentrated in vacuo. The residue was purified by column chromatography (petroleum ether/CH_2Cl_2; 2:1 v/v), and then HPLC on a Discovery C18 column (5 μm; 21.2 mm × 150 mm; Waters Corp.) eluted with 0.005% TFA in water/MeCN as mobile phase at 24 mL/min to afford **4** (t_R = 7.8−8.3 min) as a yellow oil (510 mg, 43.7%). LC-MS (ESI): *m/z* 228 $(M + H - 100)^+$, purity 100%, t_R = 2.90 min, method 2. ^{1}H NMR (300 MHz, $CDCl_3$) δ 6.13 (d, *J* = 1.5 Hz, 1H), 4.03−3.90 (m, 4H), 3.17−3.02 (m, 2H), 2.56 (t, *J* = 5.1 Hz, 2H), 2.02−1.93 (m, 2H), 1.72−1.60 (m, 2H), 1.47 (s, 9H).

*2-Fluorospiro[4,5-dihydrothieno[2,3-c]pyran-7,4′-piperidine] (**5**).* HCl (4 M; 13.6 mmol) in dioxane (3.4 mL) was added to a solution of **4** (510 mg, 1.56 mmol) in CH_2Cl_2 (3 mL). The reaction mixture was stirred for 3 h, diluted with CH_2Cl_2, and then washed with water and brine. The organic phase was dried over Na_2SO_4, filtered, and concentrated in vacuo to afford **5** as a yellow solid (278 mg, 78%). LC-MS (ESI): *m/z* 228 $(M + H)^+$, purity 93.8%, t_R = 1.99 min, method 2. ^{1}H NMR (300 MHz, $CDCl_3$) δ 6.12 (s, 1H), 3.92 (t, *J* = 5.7 Hz, 2H), 3.70 (s, 1H), 3.04−2.89 (m, 4H), 2.54 (t, *J* = 5.7 Hz, 2H), 2.02−1.97 (m, 2H), 1.72 (td, *J* = 13.2 Hz, 4.8 Hz, 2H).

*t-Butyl 3-(2-Fluorospiro[4,5-dihydrothieno[2,3-c]pyran-7,4′-piperidine]-1′-yl)propanoate (**6**).* A mixture of **5** (278 mg, 1.22 mmol), *t*-butyl acrylate (313 mg, 2.45 mmol), and TEA (367 mg, 3.67 mmol) in THF (10 mL) was heated to 65 °C, and stirred for 20 h. The reaction mixture was then cooled to rt and concentrated in vacuo. The residue was dissolved in CH_2Cl_2 and washed with 1.0 M HCl (aq) and brine. The organic layer was dried over Na_2SO_4, filtered, and concentrated in vacuo. The residue was purified by TLC (petroleum ether/EtOAc; 1:1 v/v) to afford **6** as a yellow oil (285 mg, 65.8%). LC-MS (ESI): *m/z* 356 $(M + H)^+$, purity 100%, t_R = 2.30 min, method 2. ^{1}H NMR (300 MHz, $CDCl_3$) δ 6.11 (d, *J* = 2.1 Hz, 1H), 3.91 (t, *J* = 5.4 Hz, 2H), 2.73−2.68 (m, 4H), 2.54 (t, *J* = 5.4 Hz, 2H), 2.46−2.41 (m, 4H), 2.04−1.98 (m, 2H), 1.81 (td, *J* = 13.5, 4.5 Hz, 2H), 1.46 (s, 9H).

*t-Butyl 2-[(2-Fluorophenyl)methyl]-3-(2-fluorospiro[4,5-dihydrothieno-[2,3-c]pyran-7,4′-piperidine]-1′-yl)propanoate (**7a**).* A 1 M solution of $LiN(TMS)_2$ (2.70 mmol) in THF (2.70 mL) was added dropwise to a stirred solution of **6** (318 mg, 0.90 mmol) in dry THF (20 mL) at −78 °C. The mixture was stirred for 1 h at the same temperature. 1,3-Dimethyl-tetrahydropyrimidin-2(1*H*)-one (92 mg, 0.72 mmol) in dry THF (10 mL) was added at −78 °C and stirred for 30 min. To the resulting mixture was added 1-(bromomethyl)-2-fluorobenzene (141 mg, 0.75 mmol) in dry THF (10 mL) at −78 °C. Stirring was continued for 1 h, and then the temperature was allowed to rise to 0 °C, where it was kept for 1 h. The reaction mixture was quenched with saturated NH_4Cl solution and twice extracted with EtOAc. The organic layers were combined and washed with brine, dried over Na_2SO_4, and concentrated. The residue was purified by TLC (petroleum ether/EtOAc; 8:1 v/v) to afford **7a** as a yellow oil (341 mg, 82.8%). LC-MS (ESI): *m/z* 464 $(M + H)^+$, purity 91.6%, t_R = 2.63 min, method 2. ^{1}H NMR (300 MHz, $CDCl_3$) δ 7.22−7.14 (m, 2H), 7.06−6.90 (m, 2H), 6.11 (d, *J* = 1.5 Hz,

1H), 3.90 (t, J = 5.1 Hz, 2H), 2.98−2.30 (m, 11H), 2.03−1.91 (m, 2H), 1.80−1.65 (m, 2H), 1.34 (s, 9H).

*t-Butyl 3-(2-Fluorospiro[4,5-dihydrothieno[2,3-c]pyran-7,4′-piperidine]-1′-yl)propanoate (**7b**).* Compound **6** (0.83 g, 2.33) was dissolved in dry THF (15 mL) and then cooled to −78 °C. The solution was purged with nitrogen three times. $LiN(TMS)_2$ (7.0 mL, 7.00 mmol) was added dropwise to the reaction at −78 °C, and the mixture was stirred at this temperature for 1.5 h. A solution of 1,3-dimethyl-3,4,5,6-tetrahydro-2(1*H*)-pyrimidinone (0.24 g, 1.87 mmol) in THF (5 mL) was added dropwise, and the reaction mixture was stirred for 0.5 h at −78 °C. Then a solution of 1-(bromomethyl)-2-chlorobenzene (0.72 g, 3.50 mmol) in THF (5 mL) was added. The mixture was stirred at −78 °C for 0.5 h and then at 0 °C for 0.5 h. The reaction was quenched by adding saturated aq NH_4Cl. The aqueous phase was extracted with EtOAc and then discarded. The organic phase was dried over Na_2SO_4 and concentrated. TLC (petroleum ether/EtOAc; 10:1 v/v) of the crude product gave **7b** as a solid (1.1 g, 98% yield). LC-MS (ESI): m/z: 480 $(M + H)^+$, purity 95.2%, t_R = 2.77 min, method 2. ^1H NMR (300 MHz, $CDCl_3$): δ 7.34−7.13 (m, 4H), 6.10 (s, 1H), 3.89 (t, J = 5.4 Hz, 2H), 3.05−2.29 (m, 11H), 2.01−1.90 (m, 2H), 1.80−1.65 (m, 2H), 1.34 (s, 9H).

*2-[(2-Fluorophenyl)methyl]-3-(2-fluorospiro[4,5-dihydrothieno-[2,3-c]pyran-7,4′-piperidine]-1′-yl)propanoic Acid (**8a**).* Compound **7a** (1.53 g, 3.29 mmol) was dissolved in dry CH_2Cl_2 (5 mL), and TFA (10 mL) was added. The mixture was stirred at rt for 16 h. Solvent was removed to give **8a** as a yellow oil (1.33 g). The crude material was used in subsequent steps without further purification. LC-MS (ESI): 408 $(M + H)^+$, purity 94.5%, t_R = 1.74 min, method 2.

*2-[(2-Chlorophenyl)methyl]-3-(2-fluorospiro[4,5-dihydrothieno-[2,3-c]pyran-7,4′-piperidine]-1′-yl)propanoic Acid (**8b**).* Compound **7b** (1.10 g, 2.29 mmol) was dissolved in dry CH_2Cl_2 (30 mL), and TFA (3 mL) was added. The mixture was stirred at rt for 5 h. The solvent was removed to give crude **8b** (0.97 g), which was used in subsequent steps without further purification. LC-MS (ESI): 424 $(M + H)^+$, purity 98%. t_R = 2.07 min, method 2.

*(2S)-2-[(2-Fluorophenyl)methyl]-3-(2-fluorospiro[4,5-dihydrothieno [2,3-c]pyran-7,4′-piperidine]-1′-yl)-N-isopropyl-propanamide ((**S**)-**9a**).* Di-isopropylethylamine (DIPEA; 1.29 g, 9.984 mmol) was added to a solution of **8a** (0.68 g, 1.677 mmol) in CH_2Cl_2 (20 mL), followed by 2-propanamine (0.39 g, 6.6 mmol), 1-ethyl-3(dimethylaminopropyl)-carbodiimide (EDCI) hydrochloride (0.64 g, 3.34 mmol), and 1-hydroxybenzotriazole (HOBt, 0.45 g, 3.33 mmol). The reaction mixture was stirred at rt overnight, diluted with CH_2Cl_2 (50 mL), washed with $NaHCO_3$ (aq) and brine, dried over Na_2SO_4, and evaporated to dryness. The residue was purified with TLC (CH_2Cl_2/MeOH; 25:1 v/v) to afford **9a** (0.65 mg, 88%). LC-MS ESI m/z: 449 $(M + 1)^+$, purity 98%, t_R = 5.05 min, method 2 . Chiral HPLC of **9a** on a Chiralpak AD column: (10 μm; 4.6 mm × 250 mm) eluted with hexane containing dimethylethylamine (DMEA; 0.2%) and EtOH (5%) at 1 mL/min, gave (**S**)-**9a** as a solid (268 mg): t_R = 7.34 min; mp: 139.0−139.9 °C; $[\alpha]_D$ = −7.8° (c 8.2 MeOH). ^1H NMR (300 MHz, MeOH-d_4): δ 7.23−7.18 (m, 2H), 7.08−7.00 (m, 2H), 6.22 (s, 1H), 3.92−3.83 (m, 3H), 2.89−2.65 (m, 6H), 2.54−2.32 (m, 5H), 1.99−1.94 (m, 2H), 1.81−1.70 (m, 2H), 1.18 (d, J = 6.6 Hz, 3H), 1.00 (d, J = 6.6 Hz, 3H). LC-MS (ESI): m/z 449.2 $(M + 1)^+$, purity 100%, t_R= 4.21 min, method 3. HRMS (ESI): calcd for $C_{24}H_{30}F_2N_2O_2S + H^+$, 448.2069, found 448.2073.

*(2R)-2-[(2-Fluorophenyl)methyl]-3-(2-fluorospiro[4,5-dihydrothieno [2,3-c]pyran-7,4′-piperidine]-1′-yl)-N-isopropyl-propanamide ((**R**)-**9a**).* The chiral HPLC of **9**, described above, also gave (**R**)-**9a** as a solid (271 mg): t_R = 9.26 min; mp: 137.3−139.1 °C; $[\alpha]_D$ = +8.2° (c 11.1 MeOH). ^1H NMR and LC-MS as for (**S**)-**9**, purity 100%, t_R = 4.21 min, method 3. HRMS (ESI) calcd for $C_{24}H_{30}F_2N_2O_2S + H^+$, 448.2069, found 448.2075.

*(2S)-2-[(2-Chlorophenyl)methyl]-3-(2-fluorospiro[4,5-dihydrothieno [2,3-c]pyran-7,4′-piperidine]-1′-yl)propanamide ((**S**)-**9b**).* DIPEA (0.89 g, 6.85 mmol) was added to a solution of **8b** (0.48 g, 1.14 mmol) in CH_2Cl_2 (20 mL), followed by NH_4Cl (0.18 g, 3.34 mmol), EDCI hydrochloride (0.44 g, 2.28 mmol), and HOBt (0.31 g, 2.28 mmol). The reaction mixture was stirred at rt overnight, diluted with CH_2Cl_2 (50 mL), washed with $NaHCO_3$ (aq) and brine, dried over Na_2SO_4, and evaporated to dryness. The residue was purified with TLC (CH_2Cl_2/MeOH, 5:1 v/v) to afford **9b** (250 mg, 52%). Chiral HPLC of **9b** on a Daicel AD-H column (5 μm; 21.2 mm × 250 mm) eluted at 12 mL/min with A/B (90:10 v/v) (A, 0.1% diethylamine (DEA) in hexane; B, 0.1% DEA in EtOH) with eluate monitored for absorbance at 214 nm gave (**S**)-**9b** (80 mg; t_R = 13.29 min); mp: 127.5−130.1 °C; $[\alpha]_D$ = −12.7° (c 10.0 MeOH). LC-MS (ESI): m/z: 423 $(M + H)^+$, purity: 99%, t_R = 4.59 min, method 2. ^1H NMR (300 MHz, $CDCl_3$) δ 7.64 (brs, 1H), 7.35−7.24 (m, 2H), 7.21−7.12 (m, 2H), 6.11 (d, J = 1.2 Hz, 1H), 5.25 (brs, 1H), 3.86 (t, J = 5.4 Hz, 2H), 3.35−3.26 (m, 1H), 2.89−2.61 (m, 5H), 2.54−2.44 (m, 3H), 2.41−2.36 (m, 1H), 2.24−2.12 (m,1H), 2.01−1.92 (m, 2H), 1.81−1.60 (m, 2H). HRMS (ESI): calcd for $C_{21}H_{24}ClFN_2O_2S + H^+$, 423.1304, found 423.1306.

*(2R)-2-[(2-Chlorophenyl)methyl]-3-(2-fluorospiro[4,5-dihydrothieno [2,3-c]pyran-7,4′-piperidine]-1′-yl)propanamide ((**R**)-**9b**).* The chiral HPLC of **9b** (90 mg), described above, also gave (**R**)-**9b** (t_R = 15.74 min; 90 mg); mp: 126.1−129.4 °C; $[\alpha]_D$ = +12° (c 10.0 MeOH). ^1H NMR and LC-MS, as for (**S**)-**9b**, purity 100%, t_R = 4.59 min, method 2. HRMS (ESI): calcd for $C_{21}H_{24}ClFN_2O_2S + H^+$, 423.1304, found 423.1303.

*(2S)-2-[(2-Fluorophenyl)methyl]-3-(2-fluorospiro[4,5-dihydrothieno [2,3-c]pyran-7,4′-piperidine]-1′-yl)propanamide ((**S**)-**9c**).* Di-isopropylethylamine (DIPEA) (1.41 g, 10.9 mmol) was added to a solution of crude **8a** (0.74 g, 1.82 mmol) in CH_2Cl_2 (20 mL), followed by NH_4Cl (0.29 g, 5.54 mmol), EDCI hydrochloride (0.70 g, 3.63 mmol), and HOBt (0.49 g, 3.63 mmol). The reaction mixture was stirred at rt overnight. The mixture was then diluted with CH_2Cl_2 (50 mL), washed with $NaHCO_3$ (aq) and brine, dried over Na_2SO_4, and evaporated to dryness. The crude material was purified with TLC (CH_2Cl_2/MeOH; 25:1 v/v) to afford **9c** as a light-yellow oil (0.5 g, 60% yield). Chiral HPLC of **9c** on a Daicel OJ-H column: (5 μm; 21.2 mm × 250 mm); eluted at 15 mL/min with A/B (80: 20 v/v) (A, 0.1% DEA/hexane; B, 0.1% DEA/alcohol) with eluate monitored for absorbance at 214 nm gave (**S**)-**9c** (0.230 g; t_R = 9.6 min); mp: 118.2−120.0 °C; $[\alpha]_D$ = +2.6° (c 7.09 MeOH). ^1H NMR (300 MHz, $CDCl_3$): δ 7.92 (brs, 1H), 7.25−7.14 (m, 2H), 7.10−6.95 (m, 2H), 6.10 (d, J = 1.5 Hz, 1H), 5.32 (brs, 1H), 3.88 (t, J = 5.4 Hz, 2H), 3.22 (d, J = 3.9 Hz, 1H), 2.82−2.58 (m, 5H), 2.57−2.44 (m, 3H), 2.36 (d, J = 3.0 Hz, 1H), 2.17 (t, J = 12.0 Hz, 1H), 2.01 (d, J = 13.5 Hz, 2H), 1.82−1.55 (m, 2H). LC-MS (ESI): m/z: 407 $(M + 1)^+$, purity: 99%, t_R = 4.34 min, method 2. HRMS (ESI): calcd for $C_{21}H_{24}F_2N_2O_2S + H^+$, 407.1599; found, 407.1602.

*(2R)-2-[(2-Fluorophenyl)methyl]-3-(2-fluorospiro[4,5-dihydrothieno [2,3-c]pyran-7,4′-piperidine]-1′-yl)propanamide ((**R**)-**9c**).* The chiral HPLC of **9c**, described above, also gave (**R**)-**9c** (0.21 g; t_R = 8.2 min); mp: 117.4−119.9 °C; $[\alpha]_D$ = −2.9° (c 7.29 MeOH). ^1H NMR and LC-MS (ESI) as for (**S**)-**9c**; purity 100%, t_R = 4.3 min, method 2. HRMS (ESI): calcd for $C_{21}H_{24}F_2N_2O_2S + H^+$, 407.1599, found 407.1600.

*(2S)-2-[(2-Fluorophenyl)methyl]-3-(2-fluorospiro[4,5-dihydro-thieno[2,3-c]pyran-7,4′-piperidine]-1′-yl)-N-isopropyl-N-methyl-propanamide ((**S**)-**10a**).* DIPEA (0.74 g, 5.73 mmol) was added to a solution of **8a** (0.75 g, 1.84 mmol) in CH_2Cl_2 (50 mL), followed by *N*-methyl 2-propanamine, (0.280 g, 3.83 mmol), EDCI hydrochloride (0.71 g, 3.7 mmol), and HOBt (0.50 g, 3.70 mmol). The reaction mixture was stirred at rt overnight, diluted with CH_2Cl_2 (30 mL), washed with $NaHCO_3$ (aq) and brine, dried over Na_2SO_4, and evaporated to dryness. The residue was purified by chromatography to afford **10a** as an oil (0.654 g, 76.8%). Chiral HPLC on a Chiralpak AD column (10 μm; 4.6 mm × 250 mm) eluted with 10% hexane in 0.2% DMEA in EtOH at 1 mL/min gave (**S**)-**10a** as an oil (170 mg; t_R = 5.03 min); $[\alpha]_D$ = −16.5° (c 7.9 MeOH). ^1H NMR (DMSO-d_6,

300 MHz) δ (amide rotamers are observed) 7.27−7.14 (m, 2H), 7.11−7.05 (m, 2H), 6.44 (s, 1H), 4.64 and 4.03 (d J = 6 Hz, 1H), 3.83 (m, 2H), 3.25 (m, 1H), 2.83 (m, 1H), 2.59 (s, 3H), 2.72−2.23 (m, 9H), 1.92−1.81 (m, 2H), 1.61−1.52 (m, 2H), 1.15 and 0.99 (dd, J = 6 Hz, 3H), 0.85 and 0.75 (dd, J = 6 Hz, 3H). LC-MS (ESI): m/z 463.2 (M + H)$^+$, purity 100%, t_R = 4.74, method 3. HRMS (ESI): calcd for $C_{25}H_{32}F_2N_2O_2S$ + H^+, 463.2225, found 463.2228.

*(2R)-2-[(2-Fluorophenyl)methyl]-3-(2-fluorospiro[4,5-dihydrothieno [2,3-c]pyran-7,4′-piperidine]-1′-yl)-N-isopropyl-N-methyl-propanamide ((**R**)-**10a**).* The chiral HPLC of **10a** described above also gave (***R***)-**10a** as an oil (191 mg; t_R = 6.40 min); $[\alpha]_D$ = +16.0° (c 7.9 MeOH). ^{1}H NMR and LC-MS (ESI) as for (***S***)-**10a**: purity 100%, t_R = 4.75 min, method 3. HRMS (ESI): calcd for $C_{25}H_{32}F_2N_2O_2S$ + H^+, 463.2225, found 463.2227.

*(2S)-2-[(2-Chlorophenyl)methyl]-3-(2-fluorospiro[4,5-dihydrothieno [2,3-c]pyran-7,4′-piperidine]-1′-yl)-N-methyl-propanamide ((**S**)-**10b**).* DIPEA (0.89 g, 6.85 mmol) was added to a solution of **8b** (0.48 g, 1.14 mmol) in CH_2Cl_2 (20 mL), followed by methanamine hydrochloride (0.23 g, 3.43 mmol), EDCI hydrochloride (0.44 g, 2.28 mmol), and HOBt (0.31 g, 2.28 mmol). The reaction mixture was stirred at rt overnight, diluted with CH_2Cl_2 (50 mL), washed with $NaHCO_3$ (aq) and brine, dried over Na_2SO_4, and evaporated to dryness. The crude material was purified with TLC (CH_2Cl_2/MeOH; 25:1 v/v) to afford **10c** as an off-white solid (0.37 g, 75% yield). Chiral HPLC of **10b** on a Daicel AD-H column (5 μm; 21.2 mm × 250 mm) eluted with A/B (90:10 v/v) (A, 0.1% DEA in hexane; B, 0.1% DEA in EtOH) at 12 mL/min with eluate monitored for absorbance at 214 nm gave (***S***)-**10b** (100 mg, t_R = 5.63 min); mp: 161.1−165.4 °C; $[\alpha]_D$ = −23.2° (c 6.67 MeOH). ^{1}H NMR (300 MHz, $CDCl_3$): δ 7.36−7.32 (m, 1H), 7.30−7.23 (m, 1H), 7.21−7.11 (m, 2H), 6.12 (d, J = 1.5 Hz, 1H), 3.88 (t, J = 5.7 Hz, 2H), 3.26 (d, J = 5.7 Hz, 1H), 2.88−2.61 (m, 8H), 2.24 (t, J = 11.7 Hz, 1H), 2.04−1.94 (m, 2H), 1.82−1.66 (m, 3H). LC-MS (ESI): m/z: 437(M + 1)$^+$, purity 100%, t_R = 4.38 min, method 2. HRMS (ESI): calcd for $C_{22}H_{26}ClFN_2O_2S$ + H^+, 437.1460, found 437.1464.

*(2R)-2-[(2-Chlorophenyl)methyl]-3-(2-fluorospiro[4,5-dihydrothieno [2,3-c]pyran-7,4′-piperidine]-1′-yl)-N-methyl-propanamide ((**R**)-**10b**).* The chiral HPLC of **10b**, described above also gave (***R***)-**10b** (120 mg, t_R = 7.93 min); mp: 165.6−167.8 °C; $[\alpha]_D$ = +24° (c 10.0 MeOH). ^{1}H NMR and LC-MS as for (***S***)-**10b**: purity 100%, t_R = 4.38 min, method 2. HRMS (ESI): calcd for $C_{22}H_{26}ClFN_2O_2S$ + H^+, 437.1460, found, 437.1468.

*(2S)-2-[(2-Fluorophenyl)methyl]-3-(2-fluorospiro[4,5-dihydrothieno [2,3-c]pyran-7,4′-piperidine]-1′-yl)-N-methyl-propanamide ((**S**)-**10c**).* DIPEA (1.37 g, 10.6 mmol) was added to a solution of **8a** (0.54 g, 1.33 mmol) in CH_2Cl_2 (20 mL), followed by methanamine hydrochloride (0.27 g, 4.00 mmol), EDCI hydrochloride (0.51 g, 2.66 mmol), and HOBt (0.41 g, 2.68 mmol). The reaction mixture was stirred at rt overnight, diluted with CH_2Cl_2 (50 mL), washed with $NaHCO_3$ (aq) and brine, dried over Na_2SO_4, and evaporated to dryness. The residue was purified with TLC (CH_2Cl_2/MeOH; 25:1 v/v) to afford **10c** as an off-white solid (0.41 g, 73% yield). Chiral HPLC of **10c** on a Daicel AD-H column: (5 μm; 21.2 mm × 250 mm) eluted at 15 mL/min with A/B (90:10 v/v) (A, 0.1% DEA/hexane; B, 0.1% DEA/alcohol) with eluate monitored for absorbance at 214 nm gave (***S***)-**10c** as a white solid (0.18 g, t_R = 10.47 min); mp: 99.5−101.8 °C; $[\alpha]_D$ = −7.0° (c 10.0 MeOH). ^{1}H NMR (300 MHz, $CDCl_3$) δ 7.54 (br, s, 1H), 7.26−7.14 (m, 2H), 7.08- 6.95 (m, 2H), 6.11 (s, 1H), 3.86 (t, J = 5.4 Hz, 2H), 3.25−3.13 (m, 1H), 2.82−2.58 (m, 8H), 2.53 (t, J = 5.4 Hz, 3H), 2.47−2.09 (m, 2H), 1.99 (d, J = 14.4 Hz, 2H), 1.87−1.52 (m, 2H). LC-MS (ESI) m/z: 421 (M + 1)$^+$, purity: 100%, t_R = 4.48 min, method 2. HRMS (ESI): calcd for $C_{22}H_{26}F_2N_2O_2S$ + H^+: 421.1756; found 421.1756.

*(2R)-2-[(2-Fluorophenyl)methyl]-3-(2-fluorospiro[4,5-dihydrothieno [2,3-c]pyran-7,4′-piperidine]-1′-yl)-N-methyl-propanamide ((**R**)-**10c**).* The chiral HPLC of **10c** described above also gave (***R***)-**10c**, as a white solid (0.17 g; t_R = 16.98 min); $[\alpha]_D$ = +7.5° (c 10.0 MeOH). ^{1}H NMR and LC-MS, as for (***S***)-**10c**; purity: 100%, t_R = 4,48 min, method 2. HRMS (ESI): calcd for $C_{22}H_{26}F_2N_2O_2S$ + H^+, 421.1756, found 421.1762.

In Vitro NOP Receptor Binding. Radioligand binding assays are commonly used to determine the affinity (K_i) of a compound for binding to a particular receptor or target protein. A filtration-based [^{3}H]nociceptin binding assay was developed, based on previous assay formats,[29] with minor modifications. Assay incubations were performed in deep-well 96-well plates with [^{3}H]nociceptin (final assay concentration 0.2 nM) and 5−10 μg of membrane protein (isolated from CHO cells expressing cloned human NOP receptors) in a final volume of 0.5 mL of HEPES buffer (20 mM; pH 7.4) containing, 5 mM $MgCl_2$, 1 mM EGTA, 100 mM NaCl, and 0.1% bovine serum albumin (BSA). Incubations were performed for 60 min at rt, which had been found previously to be optimal and terminated by filtration through glass fiber filters (Wallac filtermat A; pretreated with 0.3% polyethyleneimine for 1 h) on a cell harvester (Tomtec Inc., Hamden, CT). The filters were washed thrice with 5 mL of ice-cold Tris·HCl buffer (50 mM, pH 7.4). Filtermats were then dried and imbedded with Meltilex scintillant A and the radioactivity counted in a Microbeta scintillation counter (Wallac). Specific binding was determined by displacement with 100 nM unlabeled nociceptin. Curves were plotted as the percent of specific binding and IC_{50} values were determined using a sigmoidal dose response curve with variable slope using GraphPad Prism (GraphPad Software; San Diego, CA). K_i values were calculated from the IC_{50} by the equation of Cheng and Prusoff,[30] where $K_i = IC_{50} \times (1 + (L/K_d))^{-1}$. L is free ligand concentration, and K_d is the equilibrium dissociation constant of [^{3}H]nociceptin at cloned human NOP receptors expressed in CHO cells and was separately determined from a receptor saturation study as 0.16 ± 0.02 nM (n = 4)

In Vitro Functional Blockade of NOP Receptor Agonist-Mediated G-Protein Activation−[^{35}S]GTPγS Binding. Agonist-mediated stimulation of G-protein coupled receptors results in the activation of membrane associated G$\alpha\beta\gamma$−protein heterotrimer complexes and represents the first step in the transduction of extracellular signals to the modification of intracellular pathways. The first step in activation of receptor-mediated activation of G$\alpha\beta\gamma$−proteins heterotrimer is the exchange of Gα subunit bound guanosine diphosphate (GDP) for guanosine triphosphate (GTP). The binding of GTP to the Gα subunit causes dissociation of the heterotrimer subunits, Gβ and Gγ, resulting in the modulation of several intracellular signaling cascades. Receptor-mediated G-protein activation can be measured using the nonhydrolyzable radiolabeled analogue of GTP, [^{35}S]GTPγS. Thus, the NOP receptor antagonist affinity (K_b) of test ligands was measured in membranes expressing cloned human NOP receptors with a [^{35}S]GTPγS binding assay, according to previously described protocols with minor modifications.[31,32]

Assays were conducted in deep-well 96-well plates in a 200 μL volume with the following buffer composition: 100 mM NaCl, 20 mM HEPES, 5 mM $MgCl_2$, 1 mM EDTA, 0.1% BSA, 3 μM GDP, 0.5 nM [^{35}S]GTPγS. NOP receptor membrane suspension was added at a concentration of 20 μg protein per well, and >90% receptor stimulation was achieved with 300 nM nociceptin. In this assay, nociceptin is a full agonist (99.8 ± 7.06% relative efficacy; n = 14) with an EC_{50} of 2.4 nM using a four-parameter fit. Wheat germ agglutinin-coated SPA (scintillation proxity assay) beads (Amersham, Arlington Hts., IL) were added at 1 mg per well to detect membrane-bound [^{35}S]GTPγS. Plates were sealed and incubated for 2 h at rt and then placed at 4 °C overnight to allow the SPA beads to settle. Plates were then counted for radioactivity in a Microbeta instrument (Wallac). Specific [^{35}S]GTPγS binding was determined as the difference in cpm observed in the absence and presence of 10 μM unlabeled GTPγS. Data were plotted as the percent

of specific [^{35}S]GTPγS bound from which IC_{50} values were determined using a sigmoidal dose response curve with variable slope and GraphPad Prism software (GraphPad Software). Antagonist affinity (K_b) was estimated according to Delapp et al.[31] using a modification of the equation of Cheng and Prusoff[30] where $K_b = IC_{50} \times (1 + (L/EC_{50}))^{-1}$.

Characterization of Test Ligands in Rat In Vivo. Doses of the ligands, **10a**−**c**, to be given to rats, were selected to be low but still allow accurate measurement by ex vivo LC-MS/MS. Thus, each test ligand (1 mg) was dissolved in β-cyclodextrin solution (25%; 1 mL) and then diluted to a final ligand concentration of 6 μg/mL. Each of these solutions was administered intravenously to separate groups of three or four rats, with each injection given in a volume of 0.5 mL/kg via the lateral tail vein. Rats were sacrificed at 10, 20, 40, or 60 min after ligand injection. Samples of hypothalamic and striatal tissue were dissected out, weighed, and placed in conical centrifuge tubes on ice. Four volumes (w/v) of acetonitrile containing 0.1% formic acid were added to each tube. These samples were then homogenized using an ultrasonic probe and centrifuged at 3.75*g* for 16 min. Supernatant liquid (100 μL) was diluted by adding sterile water (100−900 μL) in HPLC injection vials for subsequent LC-MS/MS analysis.

Ligands were analyzed with a model 1200 HPLC apparatus (Agilent Technologies, Palo Alto, CA) linked to an API 4000 mass spectrometer (Applied Biosystems, Foster City, CA, USA). The liquid chromatography was performed on a C18 column (2.1 mm × 50 mm; Agilent; part no. 971700-907) eluted with 0.1% formic acid in water/acetonitrile (40:60 by vol for (*S*)-**10a** and (*S*)-**10b**, and 30:70 by vol for (*S*)-**10c**) at 0.25 mL/min (t_R = 1.7, 1.4, and 3.0 min for (*S*)-**10a**−**c**, respectively). The ligands were detected by monitoring the precursor to product ion transitions, namely, 463.8 to 240.0 for (*S*)-**10a**, 438.325 to 240 for (*S*)-**10b**, and 421.154 to 239.8 for (*S*)-**10c**. Standards were prepared by adding known quantities of analyte to samples of brain tissue from nontreated rats and processed as described above. To normalize for amount of ligand injected and animal weight, the concentration of injected ligand found in brain region tissue, was transformed to %SUV (standardized uptake value), as follows:

$$\%\text{SUV} = (\text{ligand amount measured in tissue, } ng/mL / \text{injected ligand dose, } \mu g/\text{kg}) \times 100$$

Levels of ligand in hypothalamus, a region with high NOP receptor expression,[15,16] were taken to represent total ligand binding (specific and nonspecific). Striatal levels were used to represent nonspecific binding. It should be noted, however, that striatum does have low NOP receptor expression.[15,16] Thus, striatum is a pseudo receptor-null region rather than a true null region. As such, the difference between the ligand concentration measured in the total binding region, the hypothalamus, and that measured in the pseudonull region, the striatum, is an underestimation of the true NOP receptor-specific binding.

Radiochemistry. *Production of [^{11}C]Carbon Dioxide.* No-carrier-added (NCA) [^{11}C]carbon dioxide (~1.5 Ci) was produced with a PETtrace cyclotron (GE Medical Systems; Milwaukee, WI) according to the $^{14}N(p,\alpha)^{11}C$ reaction[33] by irradiation of nitrogen gas (initial pressure 160 psi; 75 mL volume) containing 1% oxygen with a proton beam (16.5 MeV, 45 μA) for 20 min.

Production of [^{11}C]Methyl Iodide. NCA [^{11}C]methyl iodide was produced from NCA [^{11}C]carbon dioxide via reduction to [^{11}C]methane followed by vapor phase iodination.[34] Thus, at the end of the proton irradiation, [^{11}C]carbon dioxide was delivered to a PETtrace MeI process module (GE Medical Systems, model KAB301-31001) inside a hot-cell through stainless tubing (OD 1/8 in, ID 1/16 in) over 2 min, trapped on molecular sieve (13X), and reduced to [^{11}C]methane over nickel at 360 °C. The [^{11}C]methane was recirculated over iodine at 720 °C to generate [^{11}C]methyl iodide, which was trapped on Porapak Q held in the recirculation path.

*Labeling of Ligands [^{11}C](**S**)-**10a**−**c** and [^{11}C](**R**)-**10c** with Carbon-11.* Each ligand was labeled with carbon-11, purified, and formulated for intravenous injection, by the following general procedure. Precursor *N*-desmethyl compound (0.5 mg) was added to a V-vial containing anhydrous DMSO (0.4 mL) and finely ground KOH (4 mg). [^{11}C]Methyl iodide, produced from a 20 min cyclotron irradiation, was released from the Porapak trap at 180 °C into a stream of nitrogen, and bubbled into the DMSO at 17 mL/min until the radioactivity in the vial maximized. The sealed mixture was then heated at 80 °C for 5 min, after which the crude reaction mixture was diluted with water (500 μL). The product was injected onto a Luna C18 column (10 μm, 10 mm × 250 mm; Phenomenex) eluted with MeCN (A)/0.1M $HCOONH_4$ (B) (70:30 v/v) at 6.5 mL/min for [^{11}C](*S*)-**10a**, with A/B (60:40 v/v) at 6.5 mL/min for [^{11}C](*S*)-**10b**, and with A/B (50:50 v/v) at 8.5 mL/min for 5 min, and then 6 mL/min, for [^{11}C](*S*)-**10c** or [^{11}C](*R*)-**10c**. Eluate was monitored for absorbance at 235 nm (Beckman 166 UV detector) and for γ-radiation with a semiconductor pin-diode detector (Bioscan). The radioligand peak (t_R = 11.20, 10.1, 11.83, and 11.83 min for [[^{11}C](*S*)-**10a**−**c** and [^{11}C](*R*)-**10c**, respectively) was collected and mobile phase removed under high vacuum at 80 °C for 1 min. Ethanol (1.5 mL) was added to dissolve the radioactive residue and the solution then diluted with saline (13.5 mL). Finally, this solution was sterilized by filtration through a sterile Millex-MP filter (Millipore Corp.) to provide radioligand ready for intravenous injection, pending HPLC analysis.

Each radioligand was analyzed for radiochemical purity, specific activity, and chemical purity with reverse phase HPLC (see Supporting Information for elution conditions and retention times). Eluates were monitored for radioactivity (pin diode detector; Bioscan) and absorbance at 235 nm. The response of the absorbance detector was calibrated for mass of radioligand to allow determination of the mass of carrier in radioligand injectate for subsequent calculation of specific radioactivity (as Ci of radioligand/μmol of carrier). Samples were injected alone for the radiochemical purity, chemical purity, and specific activity measurements and then coinjected with the reference nonradioactive compound to check for coelution and thus radiotracer identity. The identity of each product was verified by LC-MS/MS of carrier in the radioligand preparation on a Finnigan LCQ DECA instrument.

The chiral purity of [^{11}C](*S*)-**10c** was confirmed by HPLC on a Chiralpak AD column (4.6 mm × 250 mm) eluted at 2.0 mL/min with mobile phase composed of 0.1% TEA in hexane/0.1% TEA in isopropyl alcohol (96:4 v/v). [^{11}C](*S*)-**10c** and (*S*)-**10c** had retention times of 10.7 min and the enantiomer (*R*)-**10c** a retention time of 12.35 min (see Supporting Information).

Computation and Measurement of Lipophilicity. The cLogD values for ligands (*S*)-**10a**−**c** at pH 7.4 were computed with Pallas 3.7 software (CompuDrug). The log *D* value of (*S*)-**10c** (at pH 7.4) was measured with [^{11}C](*S*)-**10c** by a previously described method.[35,36] log *D* values at pH 7.4 were also measured for enantiomers of ligands **10a**−**c** by a shake flask method.

PET Experiments in Monkeys. For each scanning session, the subject monkey was immobilized with ketamine and maintained under anesthesia with 1−3% isoflurane in oxygen. An intravenous perfusion line, filled with saline (0.9% w/v), was used for bolus injection of high purity radioligand (>99.9% radiochemical purity). PET serial dynamic images of brain were obtained for up to 90 min on either an Advance (GE Medical Systems, WI) or Focus 220 (Siemens) PET camera.

Decay-corrected time−activity curves (TACs) were obtained for 16 irregular volumes of interest (VOIs) in brain (anterior cingulate, amygdala, basal frontal cortex, caudate, cerebellum hippocampus, hypothalamus, insula, lateral temporal cortex, medial temporal cortex, occipital cortex, parietal cortex, posterior cingulate, prefrontal cortex, putamen, and thalamus). Radioactivity levels in VOIs were normalized

for injected dose and monkey weight by expression as a % standardized uptake value (%SUV):

$$\%\text{SUV} = \%\ \text{injected dose per mL} \times \text{body weight in g}$$

The following scans were performed with [^{11}C](*S*)-**10c**. *Baseline scans*: Five male monkeys (9.5−12.9 kg) received brain scans at baseline after a bolus intravenous injection of radioligand (4.92−6.60 mCi; dose of carrier (*S*)-**10c**, 0.127−0.355 nmol/kg), which was given at 10 min following intravenous injection of vehicle for preblocking agent. *Preblock scans*: These were performed on the same day at about 3 h after the baseline scan in each of the five monkeys. In each case, the NOP preblock agent **1** (1 mg/kg, iv) was administered over 2 at 10 min before radioligand (4.92−6.70 mCi; dose of carrier, 0.190−0.337 nmol/kg). *Displacement scans*: One monkey (11.47 kg) was scanned with [^{11}C](*S*)-**10c** (6.20 mCi; dose of carrier (*S*)-**10c**, 0.457 nmol/kg) with displacer **1** (1 mg/kg, iv) administered over 2 at 40 min from the start of the scan. One monkey (7.2 kg) was scanned with [^{11}C](*S*)-**10c** (6.78 mCi; dose of carrier (*S*)-**10c**, 0.794 nmol/kg) with naloxone (5 mg/kg, iv) administered over **2** at 37 min from the start of the scan. Baseline and preblock scans were also performed with [^{11}C](*R*)-**10c** in one monkey (wt, 9.9 kg; dose, 6.53 and 6.72 mCi; carrier, 0.496 and 0.366 nmol/kg), [^{11}C](*S*)-**10a** in two monkeys (wt, 11.5 and 12 kg; doses, 4.78−5.93 mCi; carrier, 0.0912−0.329 nmol/kg) and [^{11}C](*S*)-**10b** in three monkeys (wt, 9.1−10.1 kg; doses, 5.33−5.92 mCi; carrier, 0.123−0.411 nmol/kg). A total of six monkeys were used for all PET experiments, with one monkey studied with each of the three radioligands (*S*)-**10a**−**c** under baseline and preblock conditions to provide a within-subject comparison.

Stability of [^{11}C](*S*)-10c in Monkey Whole Blood and Plasma in Vitro and in Buffer. [^{11}C](*S*)-**10c** was incubated for 30 min in whole monkey blood (0.5 mL). A blood sample (0.20 mL) was removed and added to water (0.3 mL) to lyse the cells. A sample (0.45 mL) was then removed, added to acetonitrile (0.7 mL), and centrifuged. Then the supernatant liquid was analyzed by reverse phase radio-HPLC on a Novapak C18 column (4 μm; 100 mm × 800 mm; Waters Corp.) housed in a radial compression module (RCM 100) and eluted with MeOH/H_2O/Et_3N (80:20:0.1 by vol) at 2.5 mL/min. The stability of [^{11}C](*S*)-**10c** to incubation in sodium phosphate buffer (0.15 M, pH 7.4) for 2.5 h at rt was also assessed by reverse phase HPLC.

Plasma Protein Binding of [^{11}C](*S*)-10c.[37] The radioligand was added to human plasma, pooled from two subjects, and placed at the top of an "Amicon" Centrifree filter unit (200 μL/unit) and filtered by ultra centrifugation at 5000*g*. Then the filtrate (50 μL), remainder of filtrate and all components of the filter units were counted for radioactivity to allow calculation of f_P. The procedure was also perfomed with plasma pooled from two monkeys. Each measurement was made thrice. Values do not take account of nonspecific binding to the filter housing.

Emergence of Radiometabolites of [^{11}C](*S*)-10c in Monkey Plasma In Vivo. During each of six PET scans (three baseline and three preblock) with [^{11}C](*S*)-**10c**, blood samples were drawn periodically from the monkey femoral artery and collected in heparin-treated Vacutainer tubes. The samples were centrifuged and the plasma separated. A sample of plasma (0.45 mL) was mixed with acetonitrile (0.7 mL) and centrifuged. The supernatant liquid was analyzed with radio-HPLC on a Novapak C18 column (4 μm; 8 mm × 100 mm) eluted at 2.5 mL/min with MeOH/H_2O/Et_2N (80:20:0.1 by vol). The time-courses for percentages of radioactivity in plasma represented by [^{11}C](*S*)-**10c** and its radiometabolites were calculated.[38]

[$^3H_3$](*S*)-10c Autoradiography. Male littermates of NOP knockout and 129/S6 wild-type mice were generated by heterozygous matings (kindly provided by Dr. John Pintar, UMDNJ, Piscataway, NJ). Animals were euthanized by decapitation, and the brains were removed and frozen on dry ice. Twelve μm frozen coronal sections from both genotypes were thaw-mounted onto chrome alum/gelatin coated slides and stored at −70 °C until used. Sections were preincubated at 25 °C for 15 min in Tris-HCl buffer (50 mM) containing, 5 mM $MgCl_2$ (Mallinckrodt), 1 mM EDTA (EM Science), and 0.1% BSA. Incubations were conducted in the same buffer containing 0.6 nM [$^3H_3$](*S*)-**10c** at 25 °C for 120 min. Nonspecific binding was determined on adjacent sections in the presence of the NOP receptor antagonist **1** at a concentration of 10 μM. Sections were washed twice on ice for 10 min in preincubation buffer containing no BSA, followed by a rinse in ice-cold distilled water. Slides were dried under a cool stream of air and opposed to a phosphorimaging plate (BAS-IP TR 2025, Fuji, Tokyo, Japan) for 4 days. The plate was scanned using a BAS-5000 imager (Fuji). Quantification was performed with MCID 7.0 imaging software (Interfocus Imaging, UK). ^{3}H-Microscales (Amersham) were exposed with the slides and used for quantification of binding.

■ ASSOCIATED CONTENT

Supporting Information. X-ray analysis of (*S*)-**9c** and (*R*)-**9a**. Supplementary PET scans of [^{11}C](*S*)-**10c** and [^{11}C](*R*)-**9a**. HPLC of separation of [^{11}C]-(*S*)-**10c**. HPLC analyses of radioligands [^{11}C](*S*)-**10a**−**c**. This material is available free of charge via the Internet at http://pubs.acs.org.

■ AUTHOR INFORMATION

Corresponding Author

*Phone: 301 594 5986. Fax: 301 480 5112. E-mail: pikev@mail.nih.gov.

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■ ABBREVIATIONS USED

AUC, area-under-the-curve; Boc, *t*-butyloxycarbonyl; BSA, bovine serum albumin; DEA, diethylamine; DIPEA, di-isopropylethylamine; DMEA, dimethylethylamine; DMSO, dimethyl sulfoxide; EDCI, 1-ethyl-3(dimethylaminopropyl)-carbodiimide; f_P, plasma free fraction; GDP, guanosine diphosphate; GTP, guanosine triphosphate; HOBt, 1-hydroxybenzotriazole; MRI, magnetic resonance imaging; NCA, no-carrier-added; NFSI, *N*-fluoro-*N*-(phenylsulfonyl)benzenesulfonamide; NOP, nociceptin/orphanin FQ peptide; PET, positron emission tomography; rt, room temperature; SPA, scintillation proxity assay; SUV, standardized uptake value; TAC, time−activity curve; TEA, triethylamine; THF, tetrahydrofuran; TFA, trifluoracetic acid; VOI, volume of interest

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